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September 2016

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http://www.inl.gov

Prepared for the U.S. Department of Energy Under DOE Idaho Operations Office Contract DE-AC07-05ID14517

INL/EXT-16-39882

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INTRODUCTION

The dialkyl-monoamide class of extactant molecules has been studied for several decades as

alternatives to tributylphosphate (TBP) in PUREX-type separations for the nuclear fuel cycle.

Their advantages over TBP, as summarized in a review by Gasparini and Grossi, [1] include

greater selectivity for uranium over fission products, higher radiolytic stability, generation of

inoffensive radiolysis products, and the elimination of phosphorous. This latter point is referred

to as adherence to the CHON principal in which reagents are composed of only carbon,

hydrogen, oxygen and nitrogen, making them incinerable and thus preferable for waste treatment

purposes.

An additional advantage of the dialkyl-monoamides over TBP is the ease of their synthetic

tunability. This feature allows for metal ion selectivity to be designed into the structure of the

monoamide. For example, although n-alkane substituted monoamides have overall higher

extraction efficiencies for U(VI), branched-chain alkane-substituted monoamides are used for

selection of U(VI) over Th(IV) and Pu(IV). Branching at the α -carbon of the alkane attached to

the carbonyl carbon was shown to enhance extraction of uranium over the tetravalent ions

including zirconium, [2] and to minimize third phase formation upon high metal loading. [3] Prabhu

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et al. reported that di-2-ethylhexyl-butyramide (DEHBA) extracts U(VI) and Pu(IV), while di-2-ethylhexyl-isobutyramide (DEHiBA) selectively extracts U(VI), although with a lower D_U than for the unbranched monoamide. Thus, for applications involving U/Pu co-extraction, DEHBA would be preferred and for applications involving selective extraction of U, DEHiBA would be preferred. The structures of DEHBA and DEHiBA are shown in Fig. 1. In the Gasparini and Grossi review, the stability of monoamides toward γ -radiolysis was reported to be relatively unaffected by the nature of the alkane groups, though Mowafy reported that unsymmetrically substituted monoamides were more susceptible to radiolytic damage.

Although it has been commonly claimed that the monoamides have good radiation stability, there are few literature reports investigating this. They involve mainly post-irradiation studies of effects on solvent extraction and attempts to identify degradation products by spectroscopic or GC methods.^[5-12] The major products are invariably reported to be the corresponding amine and organic acid, suggesting rupture of the carbonyl C-N bond. For example, Musikas^[13,14] reported that the products of di-2-ethylhexyl-hexanamide radiolysis were di-2-ethylhexyl-amine and hexanoic acid. However, the secondary amine was produced in sub-stoichiometric amounts, suggesting that it hydrolyzed in the presence of acid. Similarly, Parikh et al. [8] identified the main products of di-hexyl-octanamide radiolysis using GC/GC-MS as di-hexyl-amine and octanoic acid. Uranium and plutonium extraction and subsequent stripping were unaffected by an absorbed dose of 600 kGy. This was attributed to the innocuous, water-soluble nature of the monoamide degradation products which is in stark contrast to the degradation products of tributylphosphate (TBP), [15] highlighting one of their advantages over the conventional extractant. Similarly, di-2ethylhexyl-amine and isobutanoic acid were reported by Pathak et al. as the products of DEHiBA for irradiation in the presence of 3-4 M HNO₃, and the ability to strip the loaded solvent of uranium and plutonium was maintained even at 600 kGy absorbed dose. [9]

As the monoamides continue to be developed for potential application to the nuclear fuel cycle, research has been focused on the behavior of DEHBA and DEH*i*BA, (Fig 1). The goal of this work is to provide a rigorous examination of the radiolytic decomposition of the uranium selective DEHiBA by identifying its degradation dose constants and products using mass spectrometry and measuring the impact on the extraction behavior of uranium(VI) by the irradiated solvent.

Figure 1. Structure of *di-2-ethylhexyl-butyramide* (DEHBA), (a) and *di-2-ethylhexyl-isobutyramide* (DEH*i*BA), (b).

EXPERIMENTAL

Materials

The DEH*i*BA used in the preliminary work was prepared at Colorado School of Mines (USA) using a 1.2:1 molar ratio of *iso*-butyryl chloride diluted in chloroform, added dropwise to a mixture of the secondary amine, *bis*-2-ethylhexyl-amine, diluted in chloroform and *tri*-ethyl-amine. The dropwise addition was done in a nitrogen atmosphere at 5 °C under vigorous stirring. Once mixed, the solution was refluxed for 2 hours at 60 °C. This solution was then filtered and washed with water, 10 wt % sodium carbonate, and 1 M HCl to remove unreacted *tri*-ethyl-amine as well as residual *iso*-butyryl chloride. The solution was then dried with sodium sulfate

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overnight. To purify the product, chloroform was removed by rotary evaporation followed by

vacuum distillation. Characterization by ¹H-NMR showed the product to be 99%+ pure. Upon the

need for larger volumes for irradiations and solvent extraction, DEHiBA was purchased from

Technocom (UK).

Steady State Irradiations

A 1 M solution of the monoamide was prepared by combining the appropriate amount of weighed

DEHiBA in dodecane for irradiation. Samples were irradiated using a ⁶⁰Co Nordion Gammacell

220, at a dose rate that varied from 4.73–4.34 kGy h⁻¹ during the course of the experiments. This

dose rate was initially measured using standard Fricke procedures, [16] and then subsequently

corrected for ⁶⁰Co decay. The sample chamber temperature was approximately 30 °C. The

solutions were irradiated either as dry organic solutions, or in contact with either 0.1 M HNO₃

(stripping acidity), or 4 M HNO₃ (extraction acidity). In some cases, the samples were irradiated

in static, sealed containers, considered deaerated upon exposure to even low absorbed doses.

Some samples, as indicated in the text, were irradiated with air sparging at a rate of 1 mL min⁻¹,

as delivered from a compressed air bottle through a mass flow controller. This was done to ensure

that the samples so treated remained saturated with dissolved oxygen. Following irradiation, the

samples were split for analysis by the collaborating laboratories at CEA-Marcoule and INL.

Pulsed Irradiations

Picosecond electron pulse radiolysis/transient absorption experiments were performed at the

Brookhaven LEAF facility, as described previously.^[17] Samples were irradiated in 1.00-cm

Suprasil semimicro cuvettes sealed with Teflon stoppers. The doses per pulse for various

experiments were in the range 20-40 Gy. Time-resolved kinetics were obtained using a FND-100

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silicon diode detector and digitized using a LeCroy WaveRunner 640Zi oscilloscope (4 GHz, 8

bit). Interference filters (*10-nm bandpass) were used for the wavelength selection of the

analyzing light.

Analytical Measurements

ESI-MS analysis at CEA

The mass spectrometry measurements were recorded in positive ion mode using a Bruker

Esquire-LC quadrupole ion trap equipped with an electrospray interface. A Cole Palmer syringe

infusion pump delivered the sample at a rate of 90 µL h⁻¹ to the electrospray source. The capillary

voltage was set to -4000 V. Nitrogen was employed as the drying and nebulizing gas. The drying

gas flow rate was 5.0 L min⁻¹ and the nebulizing gas pressure was set to 5.0 psi. The source

temperature was 250 °C. Spectra were acquired over a mass/charge (m/z) range of 45-2200 with

a trap drive setting of 50.0 at various skimmer voltages. Helium gas was used as the buffer gas in

the trap to improve trapping efficiency and as a collision gas for fragmentation experiments.

Each sample was prepared by dilution in acetonitrile to a concentration of 1.0 x 10⁻⁴ M. To

favor the ionization of compounds and avoid the formation of sodiated adducts, samples

irradiated without an aqueous phase were acidified before analysis by addition of 1 drop of 1 M

nitric acid. The ESI-MS measurements were taken within a short time after sample preparation to

avoid any solvent influences on the speciation of degradation products. Species were identified

by comparison with an isotopic pattern calculated using the software DataAnalysis 4.0. Ion trap

CID was used to help identify the structure of various species through fragmentation studies.

LC-ESI-MS analysis at INL

Aliquots of the same samples were also analyzed at INL in positive ion mode, using a Dionex

(Thermo Fisher, Sunnyvale, CA) UHPLC coupled to a Bruker (Billerica, MA) micro-TOFQ-II

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electrospray ionization mass spectrometer. The mass spectrometer nitrogen drying gas flow rate

was 9 mL min⁻¹, with a nebulizer gas pressure of 5.8 psi. The mass spectrometer conditions were

a capillary voltage of -4000 V and a source temperature of 220 °C. Samples were diluted by a

factor of 10^3 in 2-propanol followed by a factor of 4×10^2 in 50 % /50 % acetonitrile/water prior

to analysis. The LC eluent was isocratic with 65% acetonitrile/35% 0.1% formic acid in water at

a flow rate of 400 uL min⁻¹. Separations were achieved using 5 uL injections through a flow-

through needle onto a Kinetex 1.3 µ C18 50 mm x 2.1 mm column (Phenomenex, Torrance, CA,

USA). Each sample was analyzed in quintuplicate. DEHiBA was quantified using a calibration

curve spanning the equivalent (accounting for dilution) of 2 M to 0.1 M DEHiBA. No standards

were available for the degradation products, so their response is the integrated LC-MS peak area.

FT-IR analysis at CEA

Organic phase samples were analyzed with a Bruker vertex 70 spectrometer equipped with an

attenuated total reflectance cell. All spectra were collected between 400–4000 cm⁻¹, using 32

scans and a resolution of 4 cm⁻¹.

Uranium Solvent Extraction

Uranium solvent extraction distribution ratios, D_U, were measured for both irradiated and non-

irradiated solutions at INL. The aqueous phase contained 1 x 10⁻⁵ M natural uranium at the

appropriate acidity. Preliminary experiments during this study showed that neither acid pre-

equilibration of the organic phase, nor various contact times over the range of 30 sec-10 min

affected the $D_{\rm U}$ value. Subsequently the extractions reported here used non-pre-equilibrated, 30-

sec, equal volume contacts at 23 \pm 2 °C. The maximum $D_{\rm U}$ value was obtained at an initial

aqueous phase acidity of 6.5 M HNO₃ and therefore all $D_{\rm U}$ values were measured at this initial

acidity. Stripping was performed using 0.1 M HNO₃ with 1-min contacts. The $D_{\rm U}$ value was

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determined as the ratio of the concentrations of uranium measured in the two phases after the

contact. These concentrations were determined by mass spectrometry using a Thermo X series 2

ICP-MS with a Teflon sample introduction system and platinum cones. During analysis of

organic solutions, 0.2 L min⁻¹ of 20 % oxygen in argon was added to the spray chamber to aid in

combustion of the organic material and to reduce build up on the cones. Organic solutions were

emulsified with Triton TX100 into the normal 1 % nitric acid solution used to dilute samples

prior to analysis. Mass balances were calculated by comparison to the initial aqueous feed

uranium concentration and averaged 98 ± 6 %.

RESULTS AND DISCUSSION

Degradation rates for monoamide radiolysis

By monitoring the concentration of the monoamide in organic solution as a function of

absorbed dose, the concentration-dependent rate of degradation (G, µmol J-1) and the

concentration-independent dose constant (d, kGy⁻¹) can be determined. It was found that the

concentration of DEHiBA decreased as absorbed dose increased, as shown in Fig 2. The rate was

slow and approximately 70% of the initial monoamide concentration remained intact even at an

absorbed dose of 1000 kGy, under all conditions of acidity and aeration. Given the small change

in concentration, the data in these plots are modeled equally well using exponential or linear fits.

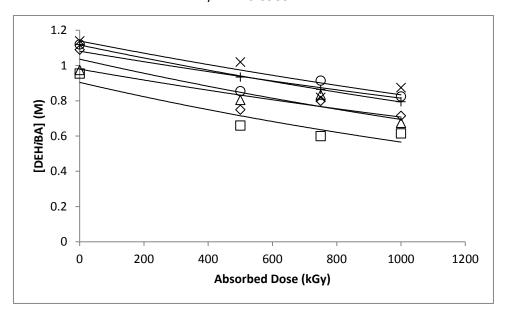


Figure 2. Change in DEH*i*BA concentration as a function of absorbed dose, for the organic phase only (+); organic phase with air sparging (circles); in contact with 0.1 M HNO₃ (diamonds); 0.1 M HNO₃ with air sparging (squares); 4 M HNO₃ (triangles); and 4 M HNO₃ with air sparging (×). Each datum is the mean of five injections with an average CI = 5.6%.

Exponential curve fitting was used to determine the dose constants (d) for monoamide degradation as the exponential constants of each equation. These values ranged from $2.8 \times 10^{-4} \text{ kGy}^{-1}$ for the aerated organic sample to $4.7 \times 10^{-4} \text{ kGy}^{-1}$ for the 0.1 M HNO_3 -contacted organic phase, aerated sample for the curves shown in Figure 2. When the complete irradiation and analyses were repeated for samples prepared from a completely separate source of DEHiBA, d ranged from $2.5 \times 10^{-4} \text{ kGy}^{-1}$ for the deaerated organic to $4.9 \times 10^{-4} \text{ kGy}^{-1}$ for the aerated organic. These values, along with the R^2 values for the exponential fits to the data, are shown in Table 1.

As can be seen in Table 1, no trends in d are found with regard to the aqueous phase or aeration condition. This suggests that all conditions had roughly the same degradation kinetics, and the range is attributable to the difficulties inherent in measuring the small concentration change that occurred across these absorbed doses. The mean value for all conditions for both trials in Table 1 is $(3.5 \pm 0.78) \times 10^{-4} \text{ kGy}^{-1}$. If the value with the very poor R^2 of 0.37 is

discounted, the mean becomes $(3.6 \pm 0.75) \times 10^{-4} \text{ kGy}^{-1}$. It is concluded that the rate constant for DEH*i*BA radiolytic degradation is essentially identical across all conditions of acidity and aeration investigated here.

These results are similar to those previously reported for diglycolamides, [17,18,19] where the aqueous phase and aeration had little or no effect on degradation rates. However, the rate of monoamide degradation is about a factor of 10 slower than for the diglycolamides. [17,18] The stability measured here seems to be of the same order of magnitude as that found for malonamides irradiated after contact with an aqueous phase of water or 1–4 M nitric acid. [20] In that study, approximately 70-80 % of the initial concentration of three different malonamides remained after an absorbed dose of 750 kGy.

In spite of the large body of literature examining TBP radiolysis, few G-values have been reported for irradiation under conditions of contact with the acidic aqueous phase, and typically the yield of the main decomposition product dibutylphosphoric acid (HDBP) is reported instead of the actual decrease in TBP concentration. These G_{HDBP} fall in the range of 0.05–0.5 μ mol J⁻¹ for alkane solutions in the absence of the aqueous phase, [15] but obviously underestimate the actual $-G_{\text{TBP}}$. Adamov reported $-G_{\text{TBP}} = 0.37 \ \mu$ mol J⁻¹ in the presence of acid. [21] If the mean dose constant reported here of 3.5 x 10^{-4} kGy⁻¹ is used to calculate a G-value [17] for the loss in initially 1 M DEHiBA, the resulting $-G_{\text{DEH}i\text{BA}} = 0.45 \ \mu$ mol J⁻¹, comparable to TBP.

Table 1. Dose constants (d, kGy^{-1}) for the degradation of DEH*i*BA (trial 1 fits to data of Fig 2; trial 2 -separate data) under various conditions of irradiation.

Condition	DEHiBA trial 1	\mathbb{R}^2	DEHiBA trial 2	\mathbb{R}^2	
Organic deaerated Organic aerated 4 M HNO ₃ deaerated	3.4 x 10 ⁻⁴ 2.8 x 10 ⁻⁴ 3.3 x 10 ⁻⁴	0.99 0.78 0.85	2.5 x 10 ⁻⁴ 4.9 x 10 ⁻⁴ 4.2 x 10 ⁻⁴	0.37 0.97 0.92	

4 M HNO ₃ aerated	3.1 x 10 ⁻⁴	0.80	3.2×10^{-4}	0.98
0.1 M HNO ₃ deaerated	4.0 x 10 ⁻⁴	0.81	2.9×10^{-4}	0.84
0.1 M HNO ₃ aerated	4.7 x 10 ⁻⁴	0.86	2.8×10^{-4}	0.96

Mean of all values = $(3.5 \pm 0.78) \times 10^{-4} \text{ kGy}^{-1}$

Organic phase analysis-FTIR

To gain information about the nature of the degradation products formed under different experimental conditions, IR analysis was performed. Figures 3 and 4 show a comparison of the sample spectrum before irradiation and after, for two conditions: irradiation with no aqueous phase present and irradiation with 4 M HNO₃ present, respectively. Figure 3 compares the impact of irradiation alone to air sparging during irradiation on monoamide degradation without an aqueous phase present. The solid line represents 1 M DEH*i*BA dissolved in dodecane before irradiation. Upon irradiation, there appears to be little impact on the nature of the degradation products in the non-air sparged sample. This could indicate that the degradation products generated by irradiation maintain similar functional groups as the parent monoamide and are thus not unambiguously observable by IR.

In the air sparged sample, the spectra are slightly different. A shoulder appears at 1175 cm⁻¹ which could represent a C-N stretch in an aliphatic amine. A new peak is also observed at 1730 cm⁻¹ and as well as a shoulder peak at 1625 cm⁻¹. The appearance of the peak at 1730 cm⁻¹ might indicate formation of a new amide or a carboxylic acid (both common degradation products previously reported for monoamides). Development of the shoulder peak at 1625 cm⁻¹ could further indicate formation of a new amide species by representing a shift in amide functionality. Other species that are commonly generated during monoamide degradation that would show C=O

stretches in this region include ketones and esters. Interestingly, based on this comparison, airsparging of the sample during irradiation changes the nature of the degradation products formed compared to irradiation under anaerobic conditions.

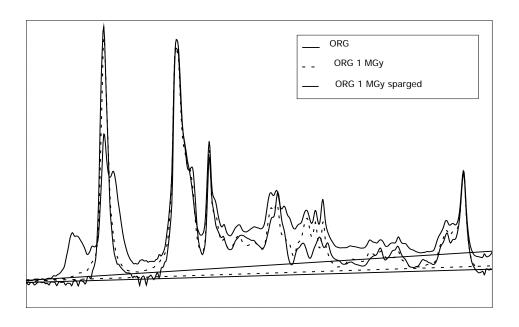
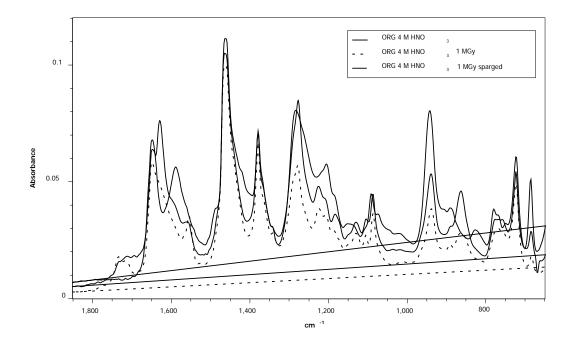


Figure 3. FTIR spectra of 1 M DEH*i*BA dissolved in dodecane (solid black line), 1 M DEH*i*BA dissolved in dodecane irradiated to 1 MGy (dashed line), 1 M DEH*i*BA dissolved in dodecane irradiated to 1 MGy with air sparge (dotted line).



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Figure 4. FTIR spectra of 1 M DEHiBA dissolved in dodecane contacted with 4 M HNO₃ (solid

black line), 1 M DEHiBA dissolved in dodecane irradiated to 1 MGy while in contact with 4 M

HNO₃ (dashed line), 1 M DEHiBA dissolved in dodecane irradiated to 1 MGy and air sparged

while in contact with 4 M HNO₃ (dotted line).

When the sample is irradiated in the presence of acid, slightly different spectra are observed,

(Fig. 4). The solid line in Fig. 4 represents 1 M DEHiBA dissolved in dodecane contacted with 4

M HNO₃. The spectrum of this sample shows a new HNO₃-monoamide peak at 1587 cm⁻¹ which

has been assigned as the C=O group bonded to HNO_{3.}[19,22,23] Other new bands appear at 1284

cm⁻¹, 940 cm⁻¹, 827 cm⁻¹, and 686 cm⁻¹, all of which are consistent with the presence of nitric acid

in the organic phase. After irradiation, in the both the air sparged and non-air sparged samples,

the same peaks appear as were observed in the air sparged sample of the organic-only irradiated

sample: 1730 cm⁻¹ and 1625 cm⁻¹. These likely represent bands from the same or similar

degradation products that were observed in the aerated, organic-only sample. Interestingly, there

is a clear decrease in the absorbance of the bands representing the C=O-HNO₃ stretch (1587 cm⁻¹

and 827 cm⁻¹) upon irradiation for both the air sparged and non-air sparged samples. This is

consistent with a decrease of the nitric acid concentration in the organic phase after irradiation.

The conclusions from this analysis are that under both oxidizing conditions of acid and air

sparging, similar degradation products are formed and these are slightly different than those that

form due to irradiation of the deaerated organic phase alone. To identify these products, the

samples were analyzed by mass spectrometry.

Organic phase analysis-ESI-MS

Mass spectra of the irradiated and non-irradiated samples show both the protonated and sodiated

parent monomer and dimer compounds at m/z = 312.2, 334.2 and m/z = 623.0, 645.5,

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respectively. Also observed in the spectra of irradiated and non-irradiated samples is a product at

m/z 242.2 corresponding to bis-2-ethylhexyl-amine (DEHA), produced by carboxyl C-N bond

rupture. It should be noted that though traces of DEHA were also found in non-irradiated

samples, likely as an impurity from synthesis, it was also clearly generated during irradiation.

Table 2 includes a summary of all degradation products found with an indication of specific

experimental conditions under which they were formed. Most products were detected regardless

of the experimental conditions though the relative yields of the degradation products were

affected by the experimental conditions.

In the low molecular weight region of the mass spectra of the irradiated solutions, several

degradation products were detected (see Table 2). The ESI-MS spectra for two of the aqueous

phase conditions (no aqueous phase, 4 M HNO₃) are shown in Fig. 5. These conditions were

shown since the spectra of the 0.1 M HNO₃ samples gave only intermediate results between these

conditions and did not provide any additional insight.

In addition to the sub-monoamide mass region, a new peak is observed at m/z 310.2 as well as

at values slightly higher than the m/z 312.2 of the monoamide. The product observed at m/z

310.2 corresponds to the abstraction of 2 hydrogen atoms from the ligand to form a C=C bond

within the monoamide framework. This product was observed regardless of acid concentration

and air sparging conditions. For samples in contact with the aqueous phase the most likely

reactive species to react by H-atom abstraction is the 'OH radical, which reacts with alkanes with

fast rate constants. The same product was found under aqueous-free conditions. In this case, the

monoamide may react with dodecane radical cations by electron transfer. This has been reported

previously for the diglycolamides.^[17] If so, the loss of H-atoms may occur through proton

elimination by the resulting monoamide radical cation (Eq. 1).

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Addition of an 'OH radical to the m/z 310.2 ion would generate the ion seen at m/z 328.0. However, this product was also observed in organic-only solution and another oxidative mechanism may account for its appearance. The product at m/z 258.2 could be produced either by carbonyl C–N bond rupture of the parent monoamide, or by H-atom abstraction from the amine N–H bond of DEHA, followed by addition of OH to generate *di-2*-ethylhexylhydroxyamine. This probably results from simple 'OH radical addition to the produced 'N radical. This product was observed only in air-sparged solutions at both acid concentrations. The product at m/z 200.2 corresponds to a C–N bond rupture resulting in the loss of one of the 2-ethylhexyl groups to produce *mono-2*-ethylhexyl-*iso*butyramide (EH*i*BA). This species was detected more at lower absorbed doses in the organic-only samples than in the 4 M HNO₃ contacted samples,

perhaps suggesting that it is susceptible to acid hydrolysis.

Table 2. Compounds observed in the organic phase as a function of experimental conditions. L stands for DEH*i*BA. N.S. stands for Non Sparged samples and S for sparged samples

m/z	Formula	Unirrad.	Org.	only	0.1 M	HNO ₃	4 M F	INO ₃
			N. S.	S.	N. S.	S.	N. S.	S.
198	$[C_{12}H_{22}NO]H^{+}$						X	
200.2	$[C_{12}H_{24}NO]H^+$		X	X	X	X	X	X
236.1						X		
240.2	$[C_{16}H_{33}N]H^{+}$		X		X		X	X
242.2	$[C_{16}H_{35}N]H^{+}$	X	X	X	X	X	X	X
254.1	$[C_{16}H_{31}NO]H^{\scriptscriptstyle +}$				X		X	X
256.0	$[C_{16}H_{33}NO]H^{\scriptscriptstyle +}$		X	X		X		
258.2	$[C_{16}H_{35}NO]H^{\scriptscriptstyle +}$					X		X
270.3				X				
284.3					X			
292.1						X		X
296.2					X			
298.3					X			
310.2	$[C_{20}H_{39}NO]H^{+}$		X	X	X	X	X	X
312.2	LH+	X	X	X	X	X	X	X
326.0	$[C_{20}H_{39}NOO]H^{\scriptscriptstyle +}$		X	X	X	X		X
328.0	$[C_{20}H_{40}NOOH]H^+$		X	X	X	X	X	
334.2	LNa^{+}	X	X	X	X	X	X	
340.2	$[C_{20}H_{37}NO_{3}]H^{+}$		X		X			
348.2						X		
350.0	LK^+	X	X	X	X	X	X	X
354.0		X	X					
364.2						X		
379.2							X	
380.1						X		
395.3								X
408.4	$[C_{28}H_{57}N]H^{+}$		X		X			
410.5	$[C_{28}H_{59}N]H^{+}$		X		X			
480.5	$[C_{32}H_{65}NO]H^{\scriptscriptstyle +}$		X	X	X			
533.5			X		X	X		
549.5	$[C_{36}H_{72}N_2O]H^+$		X		X	X		
551.7	$[C_{36}H_{74}N_2O]H^+$		X		X			
603.3						X		X
623.0	L_2H^+	X	X	X		X	X	X
645.5	L_2Na^+	X	X	X	X	X	X	X
661.1	L_2K^+	X			X	X	X	X

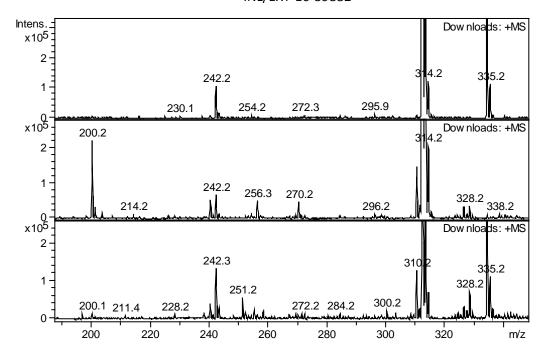


Figure 5. ESI-MS spectra of DEH*i*BA at 0 kGy (top), 1000 kGy organic phase only (middle), 1000 kGy contacted with 4 M HNO₃ (bottom). All spectra were recorded in positive ionization mode, trap drive 50, skimmer voltage 30 V, non-air sparged.

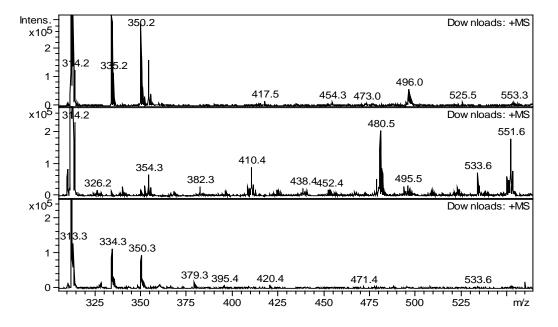


Figure 6. ESI-MS spectra of DEH*i*BA at 0 kGy (top), 750 kGy organic phase only (middle), 750 kGy contacted with 4 M HNO₃ (bottom). All spectra were recorded in positive ionization mode, trap drive 50, skimmer voltage 30 V, non-air sparged.

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For products of higher molecular weight, (Fig. 6), new species appeared at m/z 551.7, 533.5,

480.5, and 410.5. Spectra generated by MS² fragmentation suggest that m/z 551.7 is an addition

product of the monoamide with the amine DEHA. Two addition species between the dodecane or

degradation products were identified at m/z 480.5 (monoamide + dodecane) and 410.5 (DEHA +

dodecane). These high molecular weight species are likely produced by the addition reactions of

the carbon-centered radicals generated by H-atom abstractions from the corresponding reactants.

The peak at m/z 533.5 is unidentified.

Interestingly, the higher molecular weight products were observed to have the highest

intensity in the samples not contacted with an aqueous phase during irradiation (Fig. 6). For

samples irradiated in the presence of 0.1 M HNO₃, the abundance of these species was less

intense. In the spectra of the samples irradiated in the presence of 4 M HNO3 these species are

completely absent. This could indicate that the C-centered radicals were scavenged by produced

NO₃ radicals under those conditions, although it must be noted that no nitro-substituted species

were detected. Many of these nitro-, or hydroxy-substituted species may have reported to the

aqueous phase.

It is clear that the presence of acid during irradiation of the monoamide ligand impacts the

formation of higher molecular weight species. Thus, there appear to be competitive degradation

pathways for DEHiBA degradation; an acid promoted pathway; Scheme 1, and an acid

independent pathway; Scheme 2.

Scheme 1. Proposed DEH*i*BA degradation pathway in the presence of acid.

$$\begin{array}{c} & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & \\ & & & \\ &$$

Scheme 2. Proposed DEH*i*BA degradation pathway in the absence of acid.

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This conclusion correlates well with the FTIR data collected by supporting the observation

that the samples irradiated in the presence of acid resulted in new peaks in the spectrum, whereas

the organic only samples showed no difference when compared to an unirradiated monoamide

solution. Interestingly, although the product distribution is affected by contact with the acidic

aqueous phase, the overall rate of monoamide degradation is not.

Radical cation reactions with DEHiBA

The irradiation of non-acid contacted dodecane results in degradation of the diluent through

molecular ionization (e^{-} solv, $C_{12}H_{26}^{+\bullet}$) and C-H/C-C bond breakage ($C_xH_v^{\bullet}$, H^{\bullet} , H_2). Under

aerated conditions it would be expected that the carbon-centered radicals $(C_x H_y^{\bullet})$, the hydrogen

atoms (H[•]) and the solvated electrons (e⁻_{solv}) would react with dissolved oxygen to form a suite of

less-reactive peroxyl $(C_xH_yO_2^{\bullet})$ and superoxide $(HO_2^{\bullet}/O_2^{-\bullet})$ radicals. These conditions isolate

the diluent radical cation $(C_{12}H_{26}^{+\bullet})$ as the major species that could react with DEHiBA.

Therefore, in this study, we also elucidated the kinetics of this radical cation reaction with

DEHiBA in dodecane. These fast, pulsed, electron radiolysis experiments were performed as

detailed previously, [17] using 0.50 M CH₂Cl₂/aerated dodecane to isolate the radical cation with

its absorbance directly monitored at 800 nm. Typical kinetic data for the decay of this species

are seen in Figure 7. Upon addition of DEHiBA the decay becomes faster. From the analysis of

the shown decay curves the plot shown in Figure 7 (inset) is derived. The slope of this plot gives

the second-order rate constant for the reaction of $C_{12}H_{26}^{+\bullet}$ with DEHiBA as $k = (1.52 \pm 0.11) \text{ x}$

 $10^{10} \,\mathrm{M}^{-1} \,\mathrm{s}^{-1}$:

 $C_{12}H_{26}^{+\bullet} + DEHiBA \rightarrow C_{12}H_{26} + [DEHiBA]^{+\bullet}$ (1)

This effectively diffusion-controlled rate constant suggests that this reaction would be an important process in the overall organic phase degradation of this ligand, and supports the hypothesis that it may account for the appearance of m/z 310.2 when it occurs in the organic-only solution.

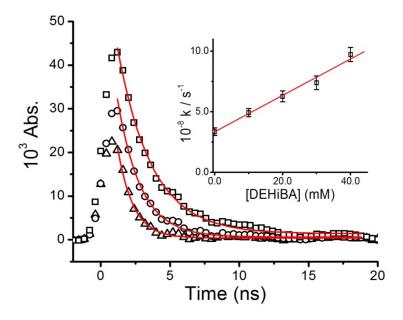


Figure 7. Transient absorption decays measured at 800 nm for dodecane radical cation reaction with $10 \, (\Box)$, $20 \, (O)$ and $40 \, (\Delta)$ mM DEHiBA at 22° C. Solid lines are fitted double-exponential decays. Inset: Rate constant determination using fast exponent fit values from kinetic traces. Slope of this line corresponds to second-order rate constant of $k = (1.52 \pm 0.11) \times 10^{10} \, \text{M}^{-1} \, \text{s}^{-1}$, $R^2 = 0.99$.

Degradation product analysis

Although the overall rate of decrease in concentration was fairly consistent under different conditions of aqueous phases and air sparging, the yields of DEHA and EHiBA varied with conditions. In the deaerated organic solution, EHiBA yields were low, with DEHA being the favored product. This is shown in Fig. 8. The very reducing conditions of the deaerated dodecane solution favored rupture of the carbonyl C–N bond (Scheme 2). The attack at this bond must be

initiated by direct effect products of dodecane; either solvated electrons, their corresponding radical cations, the 'H atom, or a combination of these species. These are also the conditions under which the higher molecular weight C-radical addition products were found. When organic-only samples were air sparged, the yield of DEHA dropped, probably due to oxygen scavenging of either solvated electrons or 'H atoms that were previously responsible for the carbonyl C–N bond rupture.

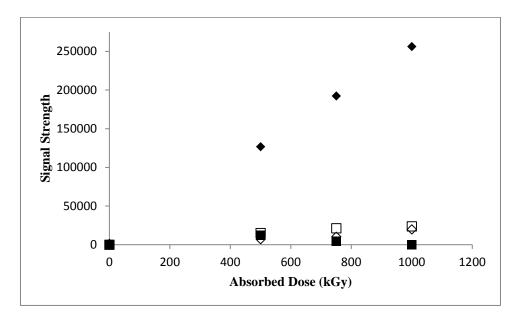


Figure 8. Yields of DEHA (m/z 242; diamonds) and EHiBA (m/z 200; squares) in irradiated 1 M DEHiBA/dodecane for deaerated (closed symbols) and air-sparged solution (open symbols). A relative standard deviation of \pm 11 % may be calculated based on quintuplicate analyses.

For the samples irradiated in the presence of 0.1 M HNO₃, shown in Fig. 9, the DEHA yield was lower, although it was still produced in highest yield in the deaerated solution. It was essentially suppressed when aerated. Since the dilute HNO₃ is expected to quantitatively scavenge radiolytically-produced electrons, the DEHA production is attributed to 'H atom reactions, and aeration further suppresses the yield through oxygen scavenging of that radical. Interestingly, aeration may have somewhat increased the yield of EH*i*BA in both the neat organic

and 0.1 M HNO₃-contacted solutions, perhaps indicating a role for peroxyl radical formation by oxygen addition to a C-centered radical for this product.

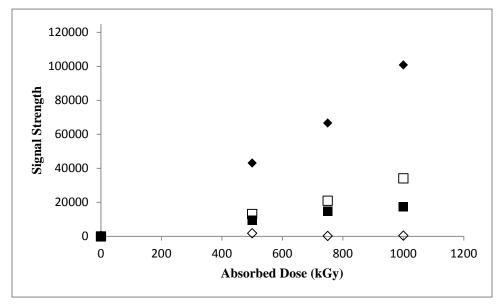


Figure 9. Yields of DEHA (m/z 242; diamonds) and EHiBA (m/z 200; squares) in the organic phase of 1 M DEHiBA/dodecane irradiated under process strip conditions of contact with 0.1 M HNO₃, for deaerated (closed symbols) and air-sparged solution (open symbols). A relative standard deviation of \pm 11 % may be calculated based on quintuplicate analyses.

The irradiations in contact with 4 M HNO₃, shown in Fig. 10, are the most oxidizing conditions studied, providing high yields of both 'OH and 'NO₃ radicals. The product yields are now reversed. Production of EH*i*BA was favored (Scheme 1), and its yield was again highest in the air-sparged solution. Thus it is concluded that DEHA is a reduction product, and that EH*i*BA is an oxidation product of DEH*i*BA radiolysis.

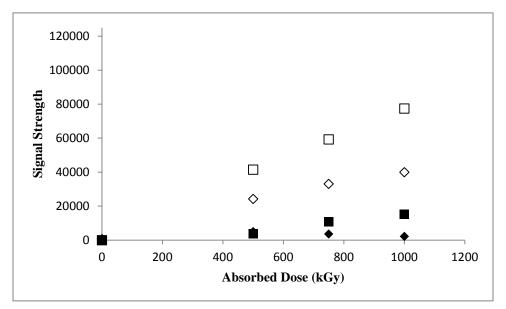


Figure 10. Yields of DEHA (m/z 242; diamonds) and EH*i*BA (m/z 200; squares) in the organic phase of 1 M DEH*i*BA/dodecane irradiated under process extraction conditions of contact with 4.0 M HNO₃, for deaerated (closed symbols) and air-sparged solution (open symbols). A relative standard deviation of \pm 11 % may be calculated based on quintuplicate analyses.

As shown in Fig. 2, the dose constants for DEH*i*BA degradation were invariant across all irradiation conditions, yet as shown in Figs. 8–10 the yields of the major products DEHA and EH*i*BA were very different. Absolute product concentrations cannot be detemined, and signal strengths may vary between mass spectrometric analytical campaigns and thus mass balance analysis for the products cannot be performed. However, from a qualitative point of view it seems clear that mass balance is not achieved based on DEHA and EH*i*BA alone, and that products with lower molecular weights than detectable here must occur, especially under oxidizing conditions. Oxygen addition to C-centered radicals is known to degrade organic compounds to carboxylic acids, ketones and aldehydes via decomposition of their peroxyl radical

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intermediates and may account for low molecular weight radiolysis product species, especially in

air sparged solution. Further, for samples irradiated in the presence of an aqueous phase,

substantial solubility of products in that phase may also occur and would be unaccounted for by

organic phase analysis.

Aqueous phase analysis

To aide in the understanding of the impact of acid during irradiation on monoamide

degradation, the aqueous phase was next analyzed to determine if degradation products could be

observed. The nature of the species identified in the aqueous phase by ESI-MS appears to depend

on nitric acid concentration. Fig. 11 shows the spectra of the 0.1 M HNO₃ irradiated DEHiBA

aqueous phase and Fig. 12 shows the 4 M HNO₃ sample. There are more species present in the 4

M HNO₃ sample than in the 0.1 M HNO₃ sample.

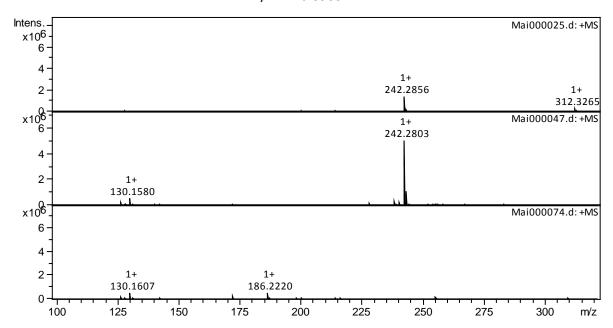


Figure 11. ESI-MS spectra of aqueous phase samples of DEH*i*BA at 0 kGy (top), 500 kGy (middle), 1000 kGy (bottom) contacted with 0.1 M HNO₃. All spectra were recorded in positive ionization mode, trap drive 50, skimmer voltage 30 V, air sparged.

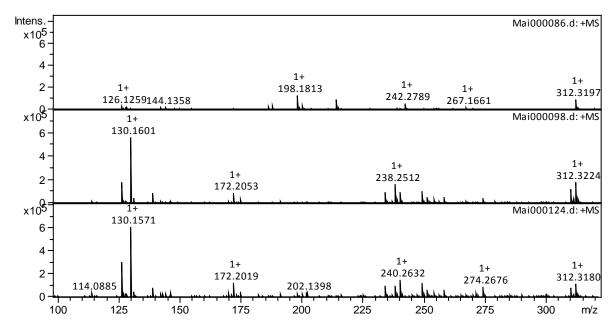


Figure 12. ESI-MS spectra of aqueous phase samples of DEH*i*BA at 0 kGy (top), 500 kGy (middle), 1000 kGy (bottom) contacted with 4 M HNO₃. All spectra were recorded in positive ionization mode, trap drive 50, skimmer voltage 30 V, air sparged.

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The products DEHA (m/z 242.2) and EHiBA (m/z 200.2) were found in both the 0.1 M and

4.0 M HNO₃ aqueous phases of the irradiated samples, with maximum signal strength in the

samples irradiated between 500-750 kGy. This is shown for samples irradiated in contact with 4

M HNO₃ in Fig. 13. A new product was detected at m/z 130.3. It has been identified as 2-

ethylhexyl-amine, indicating loss of both a butyl and one ethylhexyl group, likely produced by

continued radiolysis of either DEHA or EHiBA. The concentrations of these products decreased

at higher absorbed doses, suggesting that these species are susceptible to continued radiolysis in

the aqueous phase, and as their signals decreased, the high-intensity signal at m/z 130.3 (2-

ethylhexylamine) grew in. This signal continued to increase to a maximum at the highest

absorbed dose of 1000 kGy.

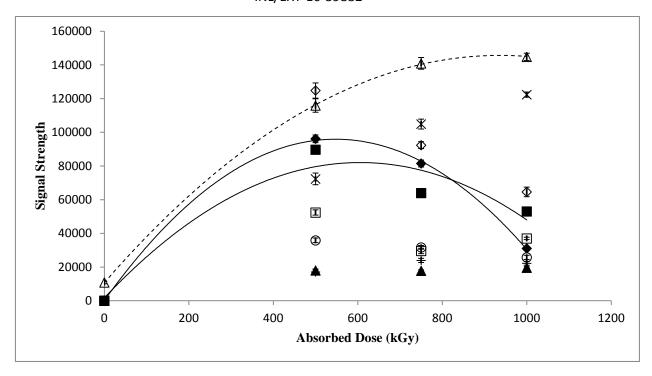


Figure 13. Yield of products in the aqueous phase found for initially 1 M DEHiBA/dodecane irradiated in contact with aerated 4 M HNO₃: DEHA (m/z 242; closed diamonds); EHiBA (m/z 200; closed squares); 2-ethylhexylamine (m/z 130, triangles); diethylhexylhydroxylamine (m/z 258, circles); (m/z 310, open diamonds); (m/z 326, open squares); (m/z 126, x); and (m/z 142, +). The parent DEHiBA also occurred at low signal strength (closed triangles). Polynomial fits given for DEHA, EHiBA and mz 310 are meant solely to guide the eye. A relative standard deviation of \pm 11 % may be calculated based on quintuplicate analyses.

An unidentified ion increases in abundance with increased dose in the 4.0 M HNO₃, air-sparged samples at m/z 142.2, and another unidentified ion was radiolytically-produced at m/z 126.2 (possibly C8H15NO). The continued degradation of the DEHA and EH*i*BA to lower molecular weight, highly oxidized species was favored under the oxidizing conditions of aeration in the presence of strong HNO₃. Small amounts of the parent monoamide were also identified in the aqueous phase. All ions detected in the aqueous phase are shown in Table 3.

Table 3. Compounds observed in the aqueous phase as a function of experimental conditions. L stands for DEH*i*BA. N.S. stands for Non Sparged samples and S for sparged samples

m/z	Formula	0.1 M HNO ₃		4 M HNO ₃	
		N. S.	S.	N. S.	S.
126.2	$[C_8H_{15}N]H^+$	X	X	X	X
130.3	$[C_8H_{19}N]H^+$	\mathbf{X}	\mathbf{X}	\mathbf{X}	\mathbf{X}
139.1				\mathbf{X}	
140.2			\mathbf{X}		
142.2		\mathbf{X}	\mathbf{X}		\mathbf{X}
144.1			\mathbf{X}		
170.1					\mathbf{X}
172.2		\mathbf{X}		\mathbf{X}	
186.2		\mathbf{X}			
198.2	$[C_{12}H_{22}NO]H^{+}$	\mathbf{X}	\mathbf{X}		\mathbf{X}
200.2	$[C_{12}H_{24}NO]H^{\scriptscriptstyle +}$	\mathbf{X}		\mathbf{X}	\mathbf{X}
202.2				\mathbf{X}	\mathbf{X}
214.2		\mathbf{X}	\mathbf{X}	\mathbf{X}	
234.1					\mathbf{X}
238.2	$[C_{16}H_{31}N]H^{+}$			\mathbf{X}	\mathbf{X}
240.2	$[C_{16}H_{33}N]H^{+}$	\mathbf{X}		\mathbf{X}	\mathbf{X}
242.2	$[C_{16}H_{35}N]H^{+}$	\mathbf{X}	\mathbf{X}		
254.2	$[C_{16}H_{31}NO]H^{\scriptscriptstyle +}$				\mathbf{X}
255.3		\mathbf{X}			
256.0	$[C_{16}H_{33}NO]H^{+}$		\mathbf{X}		
258.2	$[C_{16}H_{35}NO]H^{\scriptscriptstyle +}$				\mathbf{X}
274.2					\mathbf{X}
310.2	$[C_{20}H_{39}NO]H^{+}$		\mathbf{X}	\mathbf{X}	\mathbf{X}
312.2	LH+		\mathbf{X}	\mathbf{X}	\mathbf{X}
326.0	$[C_{20}H_{39}NOO]H^{\scriptscriptstyle +}$			\mathbf{X}	\mathbf{X}
328.0	$[C_{20}H_{40}NOOH]H^{\scriptscriptstyle +}$			\mathbf{X}	
334.2	LNa ⁺				
340.2	$[C_{20}H_{37}NO_{3}]H^{+}$			X	X

Effects of irradiation on uranium solvent extraction

The $D_{\rm U}$ for the extraction of non-irradiated DEHiBA was consistent with expectations based on literature results. ^[2,4] It can be seen in Fig. 14 that within the \pm 8% uncertainty, there was no change in uranium extraction efficiency with absorbed dose to the maximum of 1000 kGy under any irradiation condition. This is consistent with the high stability of DEHiBA under all conditions, the generation of decomposition products such as EHiBA that are likely also

extractants, and also the excess initial concentration of the ligand in comparison with the tracer metal concentration (0.01 mM). It can also be seen in Fig. 14 that the stripping D_U appears to gradually decrease with absorbed dose. This may be a result of the presence of DEH*i*BA radiolysis products in the aqueous strip solution, and the corresponding stripping of uranium by these species. Although the decreases in D_U are not large, decreased stripping distribution ratios can only be considered to be advantageous to a process separation.

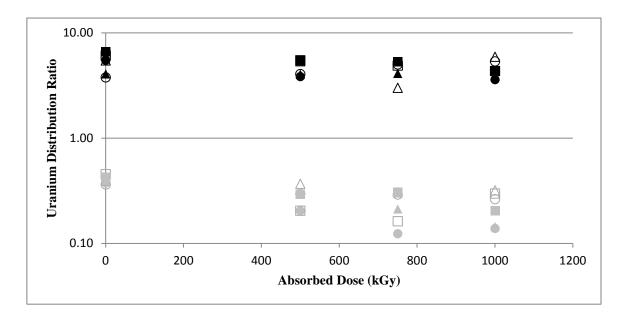


Figure 14. Uranium solvent extraction (D_U) from 6.5 M HNO₃, (black symbols) and stripping, (grey symbols) using irradiated, initially 1 M DEH*i*BA in dodecane. Irradiated organic phase only (open circles); sparged organic phase (filled circles); contact with 0.1 M HNO₃ (open triangles); sparged contact with 0.1 M HNO₃ (closed triangles); contact with 4 M HNO₃ (open squares); and sparged contact with 4 M HNO₃ (closed squares). Mean error bars of \pm 8 % omitted for clarity.

CONCLUSIONS

The radiolytic degradation rate of DEH*i*BA is similar to that of TBP and malonamides, and slow compared to the DGAs, and is unaffected by contact with an aqueous phase or aeration. However, product distributions vary with irradiation conditions. Based on these results, DEH*i*BA

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apparently undergoes degradation via two pathways: an acid promoted pathway, Scheme 1, and

an acid independent pathway, Scheme 2. It is clear that the monoamide degrades when irradiated

in the presence of an aqueous phase to form a series of lower molecular weight species generated

from the cleavage of the C-N amide bond or C-N amine bond. As this is the active site during

synthesis, it is not surprising that this is the weak point in the ligand structure. The main

degradation products appear to be DEHA and EHiBA. These species, and the smaller fragments

produced by their radioysis have increased solubility in the aqueous phase. Another product

common to all irradiation conditions was the species at m/z 310.2, which is identified as an

unsaturated derivative of DEHiBA, resulting from the loss of two H-atoms.

In contrast, when an aqueous phase is not present, higher molecular weight products are

generated via carbon radical addition reactions under the more reducing conditions. These

products have maximum abundance at 750 kGy, and then decrease with increasing absorbed

dose. Their significance to a biphasic solvent extraction process is probably inconsequential.

Solvent extraction results show that DEHiBA radiolytic degradation had little effect on

uranium distribution ratios even at absorbed doses as high as 1 MGy. The build-up of degradation

products in the aqueous phase apparently decreased stripping distribution ratios, which is not

adverse to a process application. Thus, these findings for DEHiBA are in agreement with

previous work that claimed good radiation stability and generation of inoffensive radiolysis

products for the monoamides. This, in addition to their CHON nature suggests that they will be

good candidates for the development of advanced fuel cycles. Interesting future work would

include a comparison study on the *n*-alkane monoamide DEHBA.

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